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L5 same controller	20

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Set Name side by side	Query	Hit Count	Set Name result set
DB=USPT; PLUR=YES; OP=OR			
L7	L5 same controller	20	L7
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L5	L2 same apparatus	688	L5
L4	L3 same apparatus	2	L4
L3	L2 same threshold	193	L3
L2	L1 same amplif$	26817	L2
L1	nucleic or DNA or RNA or polynucleotide or oligonucleotide	71466	L1

END OF SEARCH HISTORY

FILE 'MEDLINE' ENTERED AT 12:42:50 ON 10 JUL 2003

FILE 'BIOSIS' ENTERED AT 12:42:50 ON 10 JUL 2003
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FILE 'CAPLUS' ENTERED AT 12:42:50 ON 10 JUL 2003
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=> s nucleic or DNA or RNA or polynucleotide or oligonucleotide
L1 3487870 NUCLEIC OR DNA OR RNA OR POLYNUCLEOTIDE OR OLIGONUCLEOTIDE

=> s l1 (p)amplif?
L2 197476 L1 (P) AMPLIF?

=> s l2 (p)apparatus
L3 423 L2 (P) APPARATUS

=> s l3 (p)controller
L4 0 L3 (P) CONTROLLER

=> s l3 (p) (cycle(w)number)
L5 0 L3 (P) (CYCLE(W) NUMBER)

=> s l3(p)threshold
L6 2 L3(P) THRESHOLD

=> d bib ab l6 1-2

L6 ANSWER 1 OF 2 MEDLINE
AN 2002714647 MEDLINE
DN 22364572 PubMed ID: 12476994
TI A refined method for the detection of baculovirus occlusion bodies in forest terrestrial and aquatic habitats.
AU Ebling Peter M; Holmes Stephen B
CS Canadian Forest Service, Natural Resources Canada, 1219 Queen St E, Sault Ste Marie, Ontario, Canada P6A 2E5.. pebling@nrcan.gc.ca
SO PEST MANAGEMENT SCIENCE, (2002 Dec) 58 (12) 1216-22.
Journal code: 100898744. ISSN: 1526-498X.
CY United States
DT Journal; Article; (JOURNAL ARTICLE)
LA English
FS Priority Journals
EM 200303
ED Entered STN: 20021217
Last Updated on STN: 20030331
Entered Medline: 20030328
AB A sensitive and efficient method was developed for the detection of genetically modified and wild-type baculovirus occlusion bodies (OB) in forest terrestrial and aquatic habitats. The protocol facilitates the analysis of a large number of samples collected and frozen to maintain viral integrity. Lyophilization was used to standardize the size of both field-collected soil samples and test substrates inoculated with OBs for the determination of minimum detection **threshold**. To simulate natural conditions, terrestrial test substrates were inoculated at a standardized moisture content determined using a soil pressure plate **apparatus**. OBs, extracted from lyophilized test substrates by washing, sieving and centrifugation, were subjected to alkaline lysis and viral **DNA** isolated using a purchased **DNA** purification

kit. PCR **amplified DNA** was visualized using agarose gel electrophoresis. Minimum detection thresholds in terrestrial substrates were 10(3), 10(2), 10(2) and 10(1) OBs from 0.5 g of lyophilized L, F-H and mineral soil horizons, and 1.0 ml of leachate, respectively. Detection thresholds in aquatic substrates were 10(0) and 10(3) OBs from 1.0 ml of pond water and 1.0 g of bottom sediment, respectively.

L6 ANSWER 2 OF 2 BIOSIS COPYRIGHT 2003 BIOLOGICAL ABSTRACTS INC.
AN 2003:55966 BIOSIS
DN PREV200300055966
TI A refined method for the detection of baculovirus occlusion bodies in forest terrestrial and aquatic habitats.
AU Ebling, Peter M. (1); Holmes, Stephen B.
CS (1) Great Lakes Forestry Centre, Canadian Forest Service, 1219 Queen Street East, Sault Sainte Marie, ON, P6A 2E5, Canada: pebling@nrcan.gc.ca Canada
SO Pest Management Science, (December 2002, 2002) Vol. 58, No. 12, pp. 1216-1222. print.
ISSN: 1526-498X.
DT Article
LA English
AB A sensitive and efficient method was developed for the detection of genetically modified and wild-type baculovirus occlusion bodies (OB) in forest terrestrial and aquatic habitats. The protocol facilitates the analysis of a large number of samples collected and frozen to maintain viral integrity. Lyophilization was used to standardize the size of both field-collected soil samples and test substrates inoculated with OBs for the determination of minimum detection **threshold**. To simulate natural conditions, terrestrial test substrates were inoculated at a standardized moisture content determined using a soil pressure plate **apparatus**. OBs, extracted from lyophilized test substrates by washing, sieving and centrifugation, were subjected to alkaline lysis and viral **DNA** isolated using a purchased **DNA** purification kit. PCR **amplified DNA** was visualized using agarose gel electrophoresis. Minimum detection thresholds in terrestrial substrates were 103, 102, 102 and 101 OBs from 0.5 g of lyophilized L, F-H and mineral soil horizons, and 1.0 ml of leachate, respectively. Detection thresholds in aquatic substrates were 100 and 103 OBs from 1.0 ml of pond water and 1.0 g of bottom sediment, respectively.

=> d his

(FILE 'HOME' ENTERED AT 12:42:34 ON 10 JUL 2003)

FILE 'MEDLINE, BIOSIS, CAPLUS, EMBASE' ENTERED AT 12:42:50 ON 10 JUL 2003
L1 3487870 S NUCLEIC OR DNA OR RNA OR POLYNUCLEOTIDE OR OLIGONUCLEOTIDE
L2 197476 S L1 (P)AMPLIF?
L3 423 S L2 (P)APPARATUS
L4 0 S L3 (P)CONTROLLER
L5 0 S L3 (P) (CYCLE(W)NUMBER)
L6 2 S L3(P)THRESHOLD